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Volume 262, May 1992
Volume 31, May 1992

Pages C1181–C1188: A. Miyamoto, R. Villalobos-Molina, M. A. Kowatch, and G. S. Roth. "Altered coupling of α_1-adrenergic receptor-G protein in rat parotid during aging." Figure 5 should have appeared as follows:

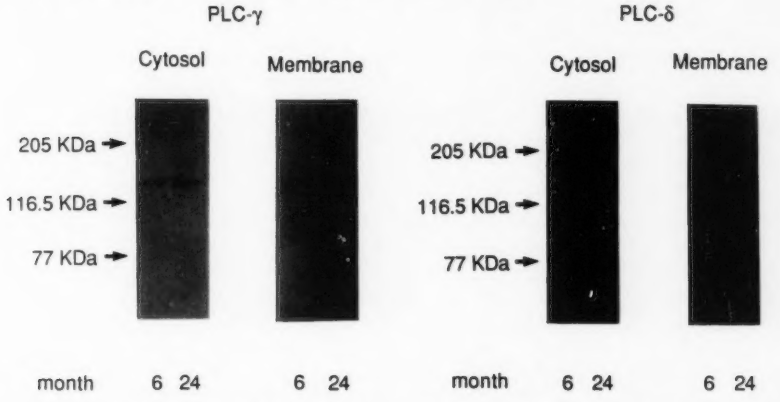

Fig. 5. Immunoblots of phosphoinositide-specific phospholipase C (PLC)-γ and PLC-δ derived from adult and old parotid cytosol and membrane preparations. Parotid cytosols and membranes from adult and old rats were subjected to SDS-PAGE followed by PLC immunoblotting using rabbit antisera and goat anti-rabbit IgG (see MATERIALS AND METHODS). Each lane contained 300 μg protein from parotid cytosol and membranes of adult and old rats. Immunolabeled PLC-γ and PLC-δ bands were identified by densitometry. The areas (AUmm) for 8 adult and 8 old cytosol were as follows: PLC-γ, 0.12 ± 0.01 and 0.10 ± 0.01, respectively, $P < 0.37$; and PLC-δ, 0.05 ± 0.00 and 0.06 ± 0.01, respectively, $P < 0.09$. The areas (AUmm) for 8 adult and 8 old membranes were as follows: PLC-γ, 0.09 ± 0.01 and 0.09 ± 0.00, respectively, $P < 0.86$; and PLC-δ, no peak found.

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Volume 262, April 1992
Volume 25, April 1992

Pages E383–E388: M. C. Michel, F. Feth, and W. Rascher. "NPY-stimulated Ca^{2+} mobilization SK-N-MC cells is enhanced after isoproterenol treatment." Page E386: line 11 on the right should read as follows: Similarly, the corresponding EC_{50} for carbachol was also not statistically significantly altered (242 ± 44 vs. 143 ± 63 μM, $P = 0.329$).

American Journal of Physiology: Gastrointestinal and Liver Physiology

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Volume 262, February 1992
Volume 6, February 1992

Pages L198–L207: Nicoletta De Marzo, David L. Sloane, Sherry Dicharry, Ella Highland, and Elliott Sigal. "Cloning and expression of an airway epithelial 12-lipoxygenase." Page L202, Fig. 3, nucleotide sequence 1321–1380 should read:

TCCTTCTGTCCCCCTGATGACCTGGCTGAC
CGGGGGCTCCTGGGAGTCAAGTCTTCTTTC

The deduced amino acid sequence for this region in Figs. 3 and 7 is correct. Also, the nucleotide sequence submitted to GenBank (accession number M62516) is correct.

Volume 263, July 1992
Volume 7, July 1992

Pages L37–L41: Gloria D. Massaro and Donald Massaro. "Formation of alveoli in rats: postnatal effect of prenatal dexamethasone." Page L39: In Table 7 the values for the number of saccules or alveoli per rat should be $\times 10^{-6}$, not $\times 10^{6}$. The corrected table appears below.

Table 7. *Dexamethasone on gestation days 17–19: average volume and number of saccules and alveoli*

Treatment	Sex	*n*	Age, days	$\tilde{v}$, $\mu m^3 \times 10^{-4}$	N, $\times 10^{-6}$
Diluent	M	3	2	19.7±3.5	1.03±0.17
Dex	M	3	2	19.8±2.5	1.04±0.23
P				NS	NS
Diluent	F	4	14	3.37±0.18	23.0±1.4
Dex	F	4	14	3.75±0.23	16.0±1.9
P				NS	<0.05

Values are means ± SE; *n*, no. of rats. Legend as in Table 4. $\tilde{v}$, volume of an average saccule or alveolus; *N*, no. of saccules or alveoli/rat.

Volume 263, August 1992
Volume 7, August 1992

Pages L219–L225: Hazel Lum, Judy L. Aschner, Patricia G. Phillips, Paul W. Fletcher, and Asrar B. Malik. "Time course of thrombin-induced increase in endothelial permeability: relationship to Ca_i^{2+} and inositol polyphosphates." Page L222: the first duration after control should be 0.5 min. The corrected table appears below.

Table 1. *F-actin content following α-thrombin treatment*

Duration of α-Thrombin (10^{-7} M) Treatment	% F-Actin/Total Protein	*n*
Control	13.98±1.4	12
0.5 min	16.85±1.7	9
1 min	25.55±1.9*	2
5 min	23.32±2.7*	10

Values are means ± SE; *n*, no. of monolayers analyzed. Triton X-100 insoluble cell extracts were prepared for SDS-PAGE analysis (see METHODS) and protein bands quantified by laser densitometry. The peak area for F-actin was normalized to total protein and reported as percent of total protein. * $P < 0.01$.

American Journal of Physiology:
Heart and Circulatory Physiology

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Volume 262, April 1992
Volume 31, April 1992

Pages H1268–H1286: M. R. Guevara and H. J. Jongsma. "Three ways of abolishing automaticity in sinoatrial node: ionic modeling and nonlinear dynamics." *Page H1284*: In the NOTE ADDED IN PROOF, the word *depolarizing* should be replaced by *hyperpolarizing*.

Volume 262, May 1992
Volume 31, May 1992

Pages H1515–H1524: A. Tajima, H. Nakata, S.-Z. Lin, V. Acuff, and J. Fenstermacher. "Differences and similarities in albumin and red blood cell flows through cerebral microvessels." *Page H1521*: right column, second paragraph, the first four lines read as follows:

If the cycling period is <10 s for the system, then all of the capillaries will be perfused and labeled within 10 s of the first appearance of RISA and radiotagged RBCs in cerebral capillaries. The time from intravenous injection to radiotracer appearance in cerebral capillaries is ~7 s in the awake rat.

Volume 262, May 1992
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Pages H1606–H1610: B. C. Simon and R. A. Cohen. "EDTA influences reactivity of isolated aorta from hypercholesterolemic rabbits." *Page H1608*: Figures 1 and 2 should read as the following.

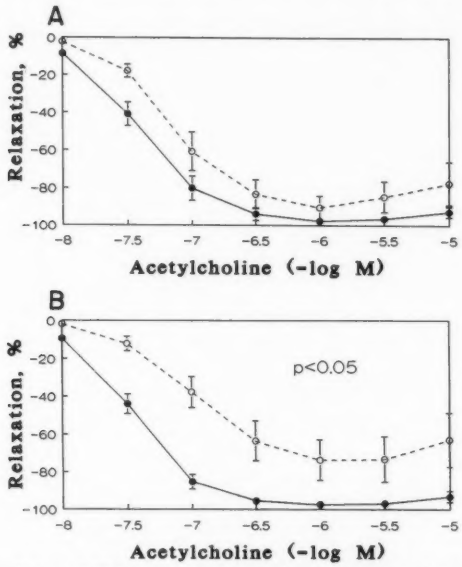

Fig. 1. Effect of EDTA on endothelium-dependent relaxation evoked by acetylcholine in aortic rings form control (filled circles, $n = 8$) and hypercholesterolemic rabbits (open circles, $n = 7$). Rings were contracted with phenylephrine, and relaxations are expressed as percentage of phenylephrine-induced tone. In absence (*B*), but not in the presence (*A*) of EDTA, relaxations of rings form hypercholesterolemic rabbits were significantly inhibited compared with those in rings from control rabbits. Comparisons were made by analysis of variance where $P <$ 0.05 was considered significant.

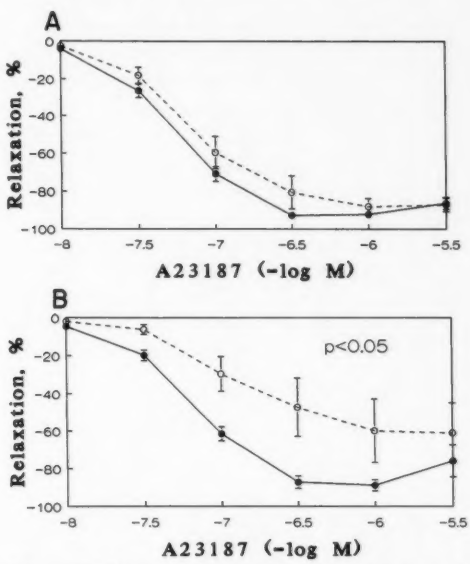

Fig. 2. Effect of EDTA on endothelium-dependent relaxation to A23187 in aortic rings from control (filled circles, $n = 8$) and hypercholesterolemic rabbits (open circles, $n = 7$). Rings were contracted with phenylephrine, and relaxations are expressed as percentage of phenylephrine-induced tone. In absence (*B*) but not in presence (*A*) of EDTA, relaxations of rings from hypercholesterolemic rabbits were significantly inhibited compared with control rabbits. Comparisons were made by analysis of variance where $P < 0.05$ was considered significant.

Volume 263, September 1992
Volume 32, September 1992

Pages H857–H865: T. C. Gillebert, S. G. De Hert, L. J. Andries, A. H. Jageneau, and D. Brutsaert. "Intracavitary ultrasound impairs left ventricular performance: presumed role of endocardial endothelium." Because of typographical errors in the numbering of the references, some of the text citations were inconsistent. Therefore this entire article is reprinted at the end of this December issue.

American Journal of Physiology: Regulatory, Integrative and Comparative Physiology

KEY TO CATEGORIES: **1.** Comparative Physiology *Regulation and Integration in:* **2.** Autonomic Physiology; **3.** Behavior; **4.** Cardiovascular Physiology; **5.** Endocrinology; **6.** Energetics; **7.** Fluids and Electrolytes; **8.** Functional Morphology; **9.** Ingestion; **10.** Periodicity; **11.** Respiration; **12.** Temperature

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Volume 262, March 1992
Volume 31, March 1992

Pages F449–F453: E. Dafnis, M. Spohn, B. Lonis, N. A. Kurtzman, and S. Sabatini. "Vanadate causes hypokalemic distal renal tubular acidosis." The value for urine anion gap in NH_4Cl-treated animals on page F450 (last sentence in first paragraph of RESULTS), as well as on page F451 (Table 2, last value in last line), should be -82 ± 7 meq/l instead of -232 ± 27 meq/l.

Pages F468–F479: F. N. Ziyadeh, J. W. Mills, and A. Kleinzeller. "Hypotonicity and cell volume regulation in shark rectal gland: role of organic osmolytes and F-actin." Page F471: in Table 1, the units for $^{86}Rb^+$ uptake should be mmol·kg dry $wt^{-1} \cdot min^{-1}$, as mentioned in MATERIALS AND METHOD (*Uptake studies*).

Volume 262, May 1992
Volume 31, May 1992

Pages F837–F842: E. Imesch, M. Moosmayer, and B. M. Anner. "Mercury weakens membrane anchoring of Na-K-ATPase." Page F839: a line of type was inadvertently dropped from the legend to Fig. 3; it should read as follows: Enhanced trypsinolysis of Na-K-ATPase α-subunit pretreated by mercury at 1 mg protein/ml. Na-K-ATPase (10 μg protein/10 μl) was incubated either in a solution stabilizing the E_1 conformation [*A*: containing (in mM) 50 NaCl, 50 KCl, 5 $MgCl_2$, 5 ATP, 30 histidine, and 1 Tris-EDTA, pH 7.2] or in a solution stabilizing the E_2 conformation (*B*: 100 mM KCl, 5 mM $MgCl_2$, 30 mM histidine, 1 mM Tris-EDTA, pH 7.2, for 30 min at 37°C) with $HgCl_2$ concentrations ranging from 10 nM to 100 μM. Trypsinolysis (30 min at 20°C) and gel electrophoresis were performed as described in EXPERIMENTAL PROCEDURES.

Advances in Physiology Education

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